

NATIONAL INFECTION PREVENTION & CONTROL MANUAL

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| | | | | Transmar Folloy | | |
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National infection prevention and control manual for England

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Version history

| Date | Version | Summary of change | | |
|-----------------|---------|--|--|--|
| 24.04.2023 | V2.5 | Link to newly published A-Z of pathogens resource added. | | |
| | | Link to newly published IPC BAF added. | | |
| | | Roles and responsibilities for staff providing care updated to include good uniform practice (bare below the elbows). | | |
| | | Section 1.2 - updated to state nail brushes should not be used. | | |
| | | Section 1.9 - updated to reflect changes to HTM 07-01. | | |
| | | Chapter 2, introduction - footnote 3 added to highlight return to prepandemic social distancing. | | |
| | | Section 2.3 - terminal cleaning of outpatient/theatre recovery amended. | | |
| | | Section 2.4 - additional recommendations on use of FRSM as source control. | | |
| | | Appendix 3 - updated to align with section 1.2. | | |
| | | Appendix 11a - recommendations on optimal patient placement and RPE use for TB updated. | | |
| 09.01.2023 V2.4 | | Hyperlinks to external documents updated. | | |
| | | Section 1.2 - additional guidance on the use of oversleeves added. Section 1.4 - amended wording of recommendations for use of sterile gloves and gowns. Section 1.8 – additional information on use of locally approved products for the management of blood and body fluid spills. Additional recommendations added to section 2.1 on cohorting of infectious patients in hospital settings. Appendix 7 updated to incorporate process for high risk body fluids (as in box 1 of appendix 9). EN standards for products used in the management of blood and body fluids spills added to the NIPCM glossary. | | |
| 07.09.2022 | V2.3 | Addition of appendices 5a and 5b and removal of previous appendix 5 (glove selection). minor change to appendix 11a to clarify precautions for varicella zoster (shingles). | | |
| 30.08.2022 | V2.2 | Minor amendments made throughout as suggested by HSE for clarity and to reflect the language used by HSE. | | |
| 21.07.2022 | V2.1 | Additional text added to appendix 11b to reflect changes in HCID status of MPX (2022 outbreak strain) and signpost to UKHSA guidance | | |
| 24.05.2022 | V2.0 | Correction to section 1.9 - safe management of waste | | |
| 14.04.2022 | V1.0 | First publication | | |

Introduction

The UK Antimicrobial five-year national action plan, published in January 2019 stated that the Scottish NIPCM will be adopted in England as national standards, to be measured by the regulators.

The NIPCM has been adapted for use within England to support and facilitate healthcare providers to demonstrate compliance with the ten criteria of the 'Health and Social Care Act 2008, Code of practice on the prevention and control of infections and related guidance (hereafter referred to as The "Code of Practice").

Aims

The NIPCM has been produced to:

- provide an evidence-based practice manual for use by all those involved in care provision in England and should be adopted as guidance in NHS settings or settings where NHS services are delivered and the principles should be applied in all care settings.
- ensure a consistent UK wide approach to infection prevention and control, however some operational and organisational details may differ across the nations.

In all non-NHS care settings, to support with health and social care integration, the content of this manual is considered best practice. The manual aims to:

- make it easy for care staff to apply effective infection prevention and control precautions
- reduce variation and optimise infection prevention and control practices across care settings in England
- improve the application of knowledge and skills in infection prevention and control
- help reduce the risk of Healthcare Associated Infection (HCAI)
- help with alignment of practice, education, monitoring, quality improvement and scrutiny.

Pathogen specific guidance is out with the remit of the NIPCM, which is not pathogen specific. Pathogen specific guidance appropriate to England is produced by other agencies for example UKHSA and can be found in the A-Z of pathogens resource https://www.england.nhs.uk/publication/national-infection-prevention-and-control/. The NIPCM outlines evidence based standard and transmission-based infection and prevention mitigations. The literature reviews that underpin and inform the practical application of the NIPCM and highlight implications for research are available at: http://www.nipcm.hps.scot.nhs.uk/resources/literature-reviews/

Audience and target groups

This manual is guidance for the NHS and as such should be applied by all NHS staff involved in patient care. Furthermore, the principles in this manual should be applied across all care settings (including acute, community and social care), complementing specific guidance produced for these settings.

Scope

Guidance contained within this manual relates to infection prevention and control practice with a primary focus on healthcare (NHS) settings, however, the principles set out are relevant to all settings where care is delivered. This manual is complemented by setting and organism specific guidance, produced by agencies such as UKHSA, and links to other relevant guidance and legislation are provided for reference.

Responsibilities for the content of this manual

NHS England will ensure that there is appropriate consultation with key stakeholders, to ensure that the NIPCM recommendations are appropriate to the system and aligned with the relevant pathogen specific guidance, legislation, and mandatory requirements for England.

Responsibilities for adopting and implementing this manual

All registered care providers must demonstrate compliance with the Health and Social Care Act 2008: Code of practice on the prevention and control of infections and related guidance which outlines ten criteria which care organisations must demonstrate compliance against.

The national IPC Board Assurance Framework (BAF) - https://www.england.nhs.uk/publication/national- infection-prevention-and-control/ is available to support organisations to effectively self-assess compliance with the code pf practice and to provide assurance in NHS settings or settings where NHS services are delivered.

Responsibilities of organisation by role

Chief executives/executive board (or equivalent) are responsible for ensuring

- systems and resources available to implement and monitor compliance with infection prevention and control (Criteria 1, Code of Practice) as specified in this guidance in all care areas; compliance monitoring includes all staff (permanent, agency and, where required, external contractors)
- there is a culture that promotes incident reporting, including near misses, while focusing on improving systemic failures and encouraging safe working practices, that is, that any workplace risk(s) are mitigated maximally for everyone. This may entail local risk assessments based on the measures as prioritised in the hierarchy of controls in the context of managing infectious agents¹
- safe systems of work, including managing the risk associated with infectious agents through the completion of risk assessments (outlined in COSHH) and approved through local governance procedures, for example Integrated Care System level. This is for the protection of all healthcare workers, patients, and visitors. This national guidance outlines the recommended principles to support local decision making within individual organisations.

Chief Operating Officers (COOs) are responsible for:

- directing the conduct of operational activities in relation to this guidance
- providing leadership, support, direction and assistance.

Directors of Infection Prevention and Control (DIPC) are responsible for ensuring:

- adoption and implementation of this guidance in accordance with local governance processes
- a workforce that is competent in IPC practice; (Criteria 6, Health and Social Care Act Code of Practice).

¹ Guidance on the hierarchy of controls is under development using the defined NIPCM methodology and will be included in the NIPCM content as a priority. Setting-specific risk assessment tools are available to support organisations in applying the HoC

Managers/employers of all services must ensure that staff:

- are aware of and have access to this guidance, including the measures required to protect themselves and their employees from infection risk
- have had instruction/education on infection prevention and control by attending events and/or completing training; (Criteria 1 and 9, Health and Social Care Act Code of Practice)
- have adequate support and resources to implement, monitor and take corrective action to comply with this guidance; and a risk assessment is undertaken and approved through local governance procedures
- who may be at high risk of complications from infection (including pregnancy) have an individual risk assessment
- who have had an occupational exposure are referred promptly to the relevant agency, e.g. GP, occupational health or accident and emergency, and understand immediate actions e.g. first aid, following an occupational exposure including process for reporting (refer to section 1.10)
- have had the required health checks, immunisations and clearance undertaken by a competent advisor (including those undertaking exposure prone procedures (EPPs); (Criteria 10, Health and Social Care Act Code of Practice)
- include infection prevention and control as an objective in their personal development plans (or equivalent) (Criteria 6, Health and Social Care Act Code of Practice)
- refer to infection prevention and control in all job descriptions.

Staff providing care must:

- show their understanding by applying the infection prevention and control principles in this guidance
- maintain competence, skills and knowledge in infection prevention and control by attending education events and/or completing training
- communicate the infection prevention and control practices to be carried out by colleagues, those being cared for, relatives and visitors, without breaching confidentiality
- have up-to-date occupational immunisations, health checks and clearance requirements as appropriate

- report to line managers, document and action any deficits in knowledge, resources, equipment and facilities or incidents that may result in transmitting infection including near misses, e.g. PPE failures
- apply the principles of good practice for uniform and workwear as set out in the NHS England uniform and workwear guidance https://www.england.nhs.uk/publication/uniforms-and-workwear-guidance-fornhs-employers/ e.g. bare below the elbows
- not provide care while at risk of transmitting infectious agents to others; if in doubt, they must consult their line manager, occupational health department, and or their infection prevention and control team (IPCT)
- inform the IPCT and local UKHSA health protection team of any outbreaks or serious incident relating to an outbreak in a timely manner and in accordance with local policies and procedures.

Infection prevention and control teams must:

- engage with staff to develop systems and processes that lead to sustainable and reliable improvements in applying infection prevention and control practices
- have suitably qualified Infection Prevention and Control staff who can provide expert advice on applying infection prevention and control in all care settings and on individual risk assessments, ensuring action is taken as required
- must maintain competence, knowledge and skills in infection prevention and control practices
- have epidemiological/surveillance systems capable of distinguishing patient case(s) requiring investigation and control.

When an organisation – eg an NHS trust – uses products or adopts practices that differ from those stated in this manual, it is responsible for ensuring safe systems of work, including the completion of a risk assessment approved through local governance procedures.

Chapter 1: Standard infection control precautions (SICPs)

Standard infection control precautions (SICPs) are to be used by all staff, in all care settings, at all times, for all patients whether infection is known to be present or not, to ensure the safety of those being cared for, staff and visitors in the care environment.

SICPs are the basic infection prevention and control measures necessary to reduce the risk of transmitting infectious agents from both recognised and unrecognised sources of infection.

Sources of (potential) infection include blood and other body fluids, secretions or excretions (excluding sweat), non-intact skin or mucous membranes and any equipment or items in the care environment that could have become contaminated.

The application of SICPs during care delivery is determined by assessing risk to and from individuals. This includes the task, level of interaction and/or the anticipated level of exposure to blood and/or other body fluids.

To protect effectively against infection risks, SICPs must be used consistently by all staff. SICPs implementation monitoring must also be ongoing to ensure compliance with safe practices and to demonstrate ongoing commitment to patient, staff and visitor safety as required by the Health and Safety Executive and the care regulators, the Care Quality Commission.

There are 10 elements of SICPs:

- 1. Patient placement/assessment of infection risk
- 2. Hand hygiene
- 3. Respiratory and cough hygiene
- 4. Personal protective equipment
- 5. Safe management of the care environment
- 6. Safe management of care equipment
- 7. Safe management of healthcare linen
- 8. Safe management of blood and body fluids
- 9. Safe disposal of waste (including sharps)
- 10. Occupational safety / managing prevention of exposure (including sharps)

1.1 Patient placement/assessment of infection risk

Patients must be promptly assessed for infection risk on arrival at the care area, e.g. inpatient/outpatient/care home, (if possible, prior to accepting a patient from another care area) and should be continuously reviewed throughout their stay.

This assessment should influence placement decisions in accordance with clinical/care need(s).

Patients who may present a cross-infection risk include those:

- with diarrhoea, vomiting, an unexplained rash, fever or respiratory symptoms
- known to have been previously positive with a Multi-drug Resistant Organism (MDRO), e.g. MRSA, CPE
- who have been an inpatient in any hospital in the UK or abroad or are a known epidemiological link to a carrier of CPE.

Further information can be found in the patient placement literature review.

1.2 Hand hygiene

Hand hygiene is considered one of the most important ways to reduce the transmission of infectious agents that cause healthcare associated infections (HCAIs).

Clinical hand-wash basins (HWBs) must:

- be used for that purpose only and not used for the disposal of other liquids
- have mixer taps, no overflow or plug and be in a good state of repair
- have wall mounted liquid soap and paper towel dispensers.

Hand hygiene facilities should include instructional posters.

Before performing hand hygiene:

- expose forearms (bare below the elbows). If disposable over-sleeves are worn for religious reasons, these must be removed and disposed of before performing hand hygiene, then replaced with a new pair*
- remove all hand and wrist jewelry. The wearing of a single, plain metal finger ring, e.g. a wedding band, is permitted but should be removed (or moved up) during hand hygiene. A religious bangle can be worn but should be moved up the forearm during hand hygiene and secured during patient care activities
- ensure fingernails are clean and short, and do not wear artificial nails or nail products

cover all cuts or abrasions with a waterproof dressing.

*refer to the NHS England uniform policy – Appendix B, for more information on the use of over- sleeves and longer sleeved uniforms.

To perform hand hygiene:

- Wash hands with non-antimicrobial liquid soap and water if:
- hands are visibly soiled or dirty
- caring for patients with vomiting or diarrhoeal illnesses
- caring for a patient with a suspected or known gastrointestinal infection, e.g. norovirus or a spore-forming organism such as Clostridioides difficile.

In all other circumstances, use alcohol-based handrubs (ABHRs) for routine hand hygiene during care.

ABHRs must be available for staff as near to the point of care as possible. Where this is not practical, personal ABHR dispensers should be used, e.g. within the community, domiciliary care, mental health units etc.

Where running water is unavailable, or hand hygiene facilities are lacking, staff may use hand wipes followed by ABHR and should wash their hands at the first opportunity.

Perform hand hygiene:

- 1. before touching a patient.
- 2. before clean or aseptic procedures.
- 3. after body fluid exposure risk.
- 4. after touching a patient; and
- 5. after touching a patient's immediate surroundings.

ALWAYS PERFORM HAND HYGIENE BEFORE PUTTING ON AND AFTER REMOVING GLOVES.

For how to wash hands, see the step-by-step guide in appendix 1 of this document. For how to hand rub, see the step-by-step guide in appendix 2 of this document.

Skin care

- dry hands thoroughly after hand washing, using disposable paper towels
- use an emollient hand cream regularly e.g. during breaks and when off duty
- do not use or provide communal tubs of hand cream in the care setting

 staff with skin problems should seek advice from occupational health or their GP and depending on their skin condition and the severity may require additional interventions or reporting.

Surgical hand antisepsis

Surgical scrubbing/rubbing (this applies to those undertaking surgical and some invasive procedures):

- perform surgical scrubbing/rubbing before donning sterile theatre garments or at other times, e.g. before inserting central vascular access devices
- remove all hand and wrist jewelry (including wedding band)
- nail brushes should not be used for surgical hand antisepsis
- nail pick (single use) can be used if nails are visibly dirty
- soft, non-abrasive, sterile (single use) sponges may be used to apply antimicrobial liquid soap to the skin if licensed for this purpose
- use an antimicrobial liquid soap licensed for surgical scrubbing or an ABHR licensed for surgical rubbing (as specified on the product label)
- ABHR can be used between surgical procedures if licensed for this use.

For surgical scrubbing (not rubbing), follow the step-by-step guide in appendix 3 of this document.

For surgical rubbing (not scrubbing), follow the step-by-step guide in appendix 4 of this document.

For hand hygiene posters/leaflet, refer to the resources section of NIPCM. Further

information can be found in the hand hygiene literature reviews:

- Hand washing, hand rubbing and indications for hand hygiene
- Hand hygiene products
- Skin Care
- Surgical hand antisepsis in the clinical setting

1.3 Respiratory and cough hygiene

Respiratory and cough hygiene is designed to minimise the risk of cross- transmission of known or suspected respiratory illness (pathogens):

- cover the nose and mouth with a disposable tissue when sneezing, coughing, wiping and blowing the nose; if unavailable use the crook of the arm
- dispose of all used tissues promptly into a waste bin
- wash hands with non-antimicrobial liquid soap and warm water after

- coughing, sneezing, using tissues, or after contact with respiratory secretions or objects contaminated by these secretions
- where there is no running water available or hand hygiene facilities are lacking, staff may use hand wipes followed by ABHR and should wash their hands at the first available opportunity
- keep contaminated hands away from the eyes nose and mouth

Staff should promote respiratory and cough hygiene helping those (e.g. elderly, children) who need assistance with this, e.g. providing patients with tissues, plastic bags for used tissues and hand hygiene facilities as necessary.

Further information can be found in cough etiquette/respiratory hygiene in the hospital setting literature review.

Personal protective equipment (PPE)

Before undertaking any procedure, staff should assess any likely exposure to blood and/or other body fluids, non-intact skin or mucous membranes and wear personal protective equipment (PPE) that protects adequately against the risks associated with the procedure. The principles of PPE use set out below are important to ensure that PPE is used correctly to ensure patient and staff safety. Avoiding overuse or inappropriate use of PPE is a key principle that ensures this is risk-based and minimizes its environmental impact. Where appropriate, consideration should be given to the environmental impact of sustainable or reusable PPE options versus single-use PPE while adhering to the principles below.

All PPE must be:

- located close to the point of use. PPE for healthcare professionals providing care in the community and domiciliary care providers must be transported in a clean receptacle
- stored to prevent contamination in a clean, dry area until required (expiry dates must be adhered to)
- single-use only unless specified by the manufacturer
- changed immediately after each patient and/or after completing a procedure or task
- disposed of after use into the correct waste stream, e.g. domestic waste, offensive (non-infectious) or clinical waste
- discarded if damaged or contaminated.

NB Reusable PPE such as non-disposable goggles/face shields/visors, must be decontaminated after each use according to manufacturer's instruction.

GLOVES must be:

- worn when exposure to blood and/or other body fluids, non-intact skin or mucous membranes is anticipated or likely.
- changed immediately after each patient and/or after completing a procedure/task even on the same patient.
- changed if a perforation or puncture is suspected
- appropriate for use, fit for purpose and well-fitting.
- never decontaminated with ABHR or soap between use
- Low risk of causing sensitisation to the wearer
- appropriate for the tasks being undertaken, taking into account the substances being handled, type and duration of contact, size and comfort of the gloves, and the task and requirement for glove robustness and sensitivity.

Sterile gloves must be worn

- when sterility is required in an operating theatre
- for some aseptic techniques e.g. insertion of central venous catheters, insertion of peripherally inserted central catheters, insertion of pulmonary artery catheters and spinal, epidural and caudal procedures.

NB Double gloving is NOT recommended for routine clinical care. However, it may be required for some exposure prone procedures, e.g. orthopaedic and gynaecological operations, when attending major trauma incidents or as part of additional precautions for high consequence infectious disease management.

Gloves are **NOT** required to carry out near patient administrative tasks, eg when using the telephone, using a computer or tablet, writing in the patient chart; giving oral medications; distributing or collecting patient dietary trays.

Further information can be found in the gloves literature review.

APRONS must be:

- worn to protect uniform or clothes when contamination is anticipated or likely, e.g. when in direct care contact with a patient.
- changed between patients and/or after completing a procedure or task.

FULL BODY GOWNS OR FLUID-RESISTANT COVERALLS must be:

 worn when there is a risk of extensive splashing of blood and/or body fluids, e.g. operating theatre, ITU

- worn when a disposable apron provides inadequate cover for the procedure or task being performed
- changed between patients and removed immediately after completing a procedure or task
- sterile when sterility is required in an operating theatre and for some asepctic techniques e.g. insertion of central venous catheters, insertion of peripherally inserted central catheters, insertion of pulmonary artery catheters and spinal, epidural and caudal procedures
- Further information can be found in the aprons/gowns literature review.

EYE OR FACE PROTECTION (INCLUDING FULL-FACE VISORS) must:

- be worn if blood and/or body fluid contamination to the eyes or face is anticipated or likely, e.g. by members of the surgical theatre team and always during aerosol generating procedures; regular corrective spectacles are not considered eye protection
- not be impeded by accessories such as piercings or false eyelashes
- not be touched when being worn.
- Further information can be found in the eye/face protection literature review.

FLUID RESISTANT SURGICAL FACE MASKS (FRSM):

Surgical face masks are required:

- as a means of source control, e.g. to protect the patient from the wearer during sterile procedures such as surgery, and
- to protect the wearer when there is a risk splashing or spraying of blood, body fluids, secretions or excretions onto the respiratory mucosa.
- As an element of PPE for droplet precautions.

FRSM must be:

- · worn (with eye protection) if a full-face visor is not available and spraying or splashing of blood, body fluids, secretions or excretions onto the respiratory mucosa (nose and mouth) is anticipated or likely (Type IIR)
- worn to protect patients from the operator as a source of infection, e.g. when performing surgical procedures or epidurals or inserting a central vascular catheter (CVC) (Type II (not classed as an FRSM) or Type IIR)

- well-fitting and fit for purpose, fully covering the mouth and nose (manufacturers' instructions must be followed to ensure effective fit and protection)
- removed or changed:
 - at the end of a procedure/task
 - if the mask's integrity is breached, e.g. from moisture build-up after extended use or from gross contamination with blood or body fluids
 - in accordance with manufacturers' specific instructions.

Further information can be found in the <u>surgical face masks literature review</u>.

FOOTWEAR must be:

- visibly clean, non-slip and well-maintained, and support and cover the entire foot to avoid contamination with blood or other body fluids or potential injury from sharps
- removed before leaving a care area where dedicated footwear is used, e.g. theatre; these areas must have a decontamination schedule with responsibility assigned.

Further information can be found in the footwear literature review

HEADWEAR

Headwear is not routinely required in clinical areas unless part of theatre attire or to prevent contamination of the environment such as in clean rooms.

HEADWEAR must be:

- worn in theatre settings and clean rooms, e.g. central decontamination unit
- well-fitting and completely cover the hair
- changed or disposed of between clinical procedures/lists or tasks and if contaminated with blood and/or body fluids
- removed before leaving the theatre or clean room
- individuals with facial hair must also cover this in areas where headwear is required, e.g. wear a snood.

NB Headwear worn for religious reasons such as turbans, kippot veil, headscarves must not compromise patient care and safety. These must be washed and/or

changed daily or immediately if contaminated and comply with additional attire in theatres etc.

Further information can be found in the headwear literature review.

For the recommended method of putting on and removing PPE, see <u>UKHSA guides</u>.

1.5 Safe management of care equipment

Care equipment is easily contaminated with blood, other body fluids, secretions, excretions and infectious agents. Consequently, it is easy to transfer infectious agents from communal care equipment during care delivery.

Care equipment is classified as either:

- single use: equipment which is used once on a single patient then discarded. This equipment must never be re-used. The packaging will carry this symbol: (2)
- single patient use: equipment which can be reused on the same patient and may require decontamination in-between use such as nebuliser masks
- reusable invasive equipment: used once then decontaminated, eg surgical instruments
- reusable non-invasive equipment: (often referred to as communal equipment) reused on more than one patient following decontamination between each use, eg commode, patient transfer trolley.

NB Needles and syringes are single use devices, they should never be used more than once or reused to draw up additional medication. Never administer medications from a single-dose vial or intravenous (IV) bag to multiple patients.

Before using any sterile equipment check that:

- the packaging is intact
- there are no obvious signs of packaging contamination
- the expiry date remains valid
- any sterility indicators are consistent with the process being completed successfully.

Decontamination of reusable non-invasive care equipment must be undertaken:

- between each use/between patients
- after blood and/or body fluid contamination
- at regular predefined intervals as part of an equipment cleaning protocol
- before inspection, servicing or repair.

If providing domiciliary care, equipment should be transported safely and decontaminated as above before leaving the patient's home.

Always adhere to COSHH risk assessments and manufacturers' guidance for use and decontamination of all care equipment.

- All reusable non-invasive care equipment must be decontaminated between patients/clients using either approved detergent wipes or detergent solution, in line with manufacturers' instructions, before being stored clean and dry.
- Decontamination protocols must include responsibility for; frequency of; and method of environmental decontamination.
- An equipment decontamination status certificate will be required if any item of equipment is being sent to a third party, eg for inspection, servicing or repair.
- Guidance should be sought from the IPCT prior to procuring, trialling or lending any reusable non-invasive equipment.
- Medical devices and other care equipment must have evidence of planned preventative maintenance programmes.

For how to decontaminate reusable non-invasive care equipment see Appendix 7.

For decontamination of surgical instruments see HTM01-01 decontamination of surgical instruments.

Further information can be found in the <u>management of patient care equipment</u> <u>literature</u> review.

1.6 Safe management of the care environment

The care environment must be:

- visibly clean, free from non-essential items and equipment to facilitate effective cleaning
- well maintained, in a good state of repair and with adequate ventilation for the clinical specialty.

Always adhere to COSHH risk assessments for product use and processes for decontamination of the care environment.

Routine cleaning

- The environment should be routinely cleaned in accordance with the National Cleaning Standards.
- Use of detergent wipes are acceptable for cleaning surfaces/frequently touched sites within the care area.
- A fresh solution of general-purpose neutral detergent in warm water is recommended for routine cleaning. This should be changed when dirty or when changing tasks.
- Routine disinfection of the environment is not recommended however, 1,000ppm available chlorine should be used routinely on sanitary fittings.
- Staff groups should be aware of their environmental cleaning schedules for their area and clear on their specific responsibilities.
- Cleaning protocols should include responsibility for, frequency of, and method of environmental decontamination.

Further information can be found in the <u>safe management of the care environment literature</u> review.

1.7 Safe management of linen

Clean linen

- Should be stored in a clean, designated area, preferably an enclosed cupboard.
- If clean linen is not stored in a cupboard, then the trolley used for storage must be designated for this purpose and completely covered with an impervious covering/or door that is able to withstand decontamination.
- Do not:
 - Rinse, shake or sort linen on removal from beds/trolleys
 - place used linen on the floor or any other surfaces eg a locker/table top
 - re-handle used linen once bagged
 - overfill laundry receptacles (not more than 2/3 full); or
 - place inappropriate items in the laundry receptacle eg used equipment/needles.

Healthcare laundry must be managed and segregated in accordance with HTM 01- 04 which categorises laundry as follows:

Used linen (previously known as soiled/fouled linen):

- Ensure a laundry receptacle is available as close as possible to the point of use for immediate linen deposit.
- Should be placed in an impermeable bag immediately on removal from the bed or before leaving a clinical department.

Infectious linen (this mainly applies to healthcare linen)

Infectious linen includes linen that has been used by a patient who is known or suspected to be infectious and/or linen that is contaminated with blood and/or other body fluids, eg faeces:

- Linen in this category must not be sorted but should be sealed in a water- soluble bag (entirely water soluble 'alginate' bag or impermeable bag with soluble seams), which is then placed in an impermeable bag immediately on removal from the bed and secured before leaving a clinical area.
- Infectious linen bags/receptacles must be tagged (eg hospital ward/care area) and dated.
- Store all used/infectious linen in a designated, safe, lockable area while awaiting uplift. Uplift schedules must be acceptable to the care area and there should be no build-up of linen receptacles.
- All linen that is deemed unfit for re-use, eg torn or heavily contaminated, should be categorised at the point of use and returned to the laundry for assessment and disposal.

Linen used during patient transfer, eg blankets, should be categorised at the point of destination.

Further information can be found in the safe management of linen literature review. For how to manage linen at care area level see Appendix 8.

Safe management of blood and body fluid spillages

Spillages of blood and other body fluids may transmit blood borne viruses. Spillages must

be treated immediately by staff trained to undertake this safely.

Responsibilities for the treatment of blood/body fluid spills must be clear within each area/care setting.

For management of blood and body fluid spillages see Appendix 9.

If an organisation locally approves a product for use in the management of blood and body fluid spills, the organisation is responsible for ensuring safe systems of work, including the completion of a risk assessment approved through local governance procedures. Organisations must confirm the efficacy and suitability of the product (i.e. that it conforms with relevant standards and is appropriate for the intended use) with the product manufacturer.

A locally approved product which conforms to: EN17126, EN13727, EN14348, EN14476, EN13697, EN14885, EN13706, EN1650, EN1276 and EN13624 may be used for the management of blood and body fluid spills.

Further information can be found in the management of blood and body fluid literature review.

Healthcare providers should ensure that any polymer gel for non-patient use (eg spill kits, controlled drug destruction, use by cleaning staff) is kept secure and away from patients. See National Patient Safety Alert - NatPSA/2019/002/NHSPS.

Safe disposal of waste (including sharps) 1.9

HTM 07:01 contains the regulatory waste management guidance for all health and care settings (NHS and non-NHS) in England and Wales including waste classification, segregation, storage, packaging, transport, treatment and disposal.

Health and Safety (Sharp Instruments in Healthcare) Regulations 2013 outline the regulatory requirements for employers and contractors in the healthcare sector in relation to the safe disposal of sharps.

Definitions

Healthcare (including clinical) waste:

Clinical Waste means waste from a healthcare activity (including veterinary healthcare) that -

- Contains viable micro-organisms or their toxins which are known or reliably believed to cause disease in humans or other living organisms
- · Contains or is contaminated with a medicine that contains a biologically active pharmaceutical agent, or
- Is a sharp, or a body fluid or other biological material (including human and animal tissue) containing or contaminated with a dangerous substance within the meaning of Regulation (EC) No 1272/2008 of the European Parliament and of the Council on classification, labelling and packaging of substances and mixtures, as amended from time to time.

for example, if a patient is known or suspected to be infected, or colonized, by an infectious agent. Clinical judgement should be applied in the assessment of waste and should consider the infection status of a patient and the item of waste produced.

Offensive Waste is waste that:

- is not clinical waste
- is not infectious, but may contain body fluids, secretions or excretions
- is non-hazardous, and
- falls within the description of code 18 01 04 if from healthcare or 20 01 99 if from municipal sources. .

Hazardous waste:

Waste that is harmful to human health or the natural environment; the Hazardous Waste (England) Regulations 2005 Annex 1 specifically lists items that are considered hazardous waste.

Table 1: Categories of waste and segregation at source

| Offensive (non-infectious) | Yellow bag with black stripe (tiger) bag | Energy from waste, landfill or other permitted processes |
|--|---|---|
| Clinical waste (infection only) | UN approved orange bag, UN approved box or sharps container | For alternative treatment |
| Healthcare waste contaminated with non- hazardous pharmaceuticals or chemicals) | UN approved yellow bag, UN approved box or sharps container | For incineration or other permitted process |
| Waste contaminated with cytotoxic or cytostatic medication | UN approved purple bag, UN approved box or sharps container | For incineration |
| Non-hazardous pharmaceuticals (no sharps) | Blue box/container | For incineration or other permitted process |
| Anatomical waste/full blood bag and blood preserves | UN approved red lidded container | For incineration only. |
| Domestic | Black/clear bags | Energy from Waste, recovery or landfill |
| Recycling | Clear, green or other colour bag | Recycling ` |

Safe waste disposal at care area level:

Always dispose of waste:

- immediately and as close to the point of use as possible; and
- into the correct segregated colour coded waste bag or rigid container or sharps box if a sharp.
- Liquid waste, e.g. suction canister must be rendered safe by adding a polymer gel or compound to the container prior to placing in an orange lidded leak proof bin.
- waste bags must be no more than 2/3 full and no more than the UN approved weight and must be securely fastened using a plastic tie or secure knot using a

- 'swan neck' to close. Waste must be traceable back to ward/care area or department, this may be achieved by writing on bags (prior to use), attaching sticky labels or unique numbered tags with the post code on them.
- store all waste in a designated, safe, lockable area while awaiting uplift. Uplift schedules must be acceptable to the care area and there should be no build-up of waste receptacles.
- local guidance on management of waste at care level, eg domiciliary settings should be followed.

Sharps containers (for safety devices, refer to section 1.10)

Sharps containers must:

- have a handle (small community boxes do not require a handle) and temporary closure mechanism, employed when box is not in use
- be disposed of when the manufacturers' fill line is reached
- be labelled with point of origin and date of assembly and disposal. Where reusable sharps containers are used, organisations must have a protocol in place to assure themselves of safe use and reprocessing.

Further information can be found in the HTM 07-01 https://www.england.nhs.uk/publication/management-and-disposal-of-healthcare-waste-htm-07-01/

1.10 Occupational safety: prevention of exposure (including sharps injuries)

The Health and Safety (Sharp Instruments in Healthcare) Regulations 2013 outline the regulatory requirements for employers and contractors in the healthcare sector in relation to: arrangements for the safe use and disposal of sharps; provision of information and training to employees; investigations and actions required in response to work related sharps injuries.

There is a potential risk of transmission of a Blood Borne Virus (BBV) from a significant occupational exposure and staff must understand the actions they should take when a significant occupational exposure incident takes place. There is a legal requirement to report all sharps injuries and near misses to line managers/employers.

A significant occupational exposure is:

- a percutaneous injury eq injuries from needles, instruments, bone fragments, or bites which break the skin; and/or
- exposure of broken skin (abrasions, cuts, eczema, etc.); and/or
- exposure of mucous membranes including the eye from splashing of blood or other high risk body fluids.

For the management of an occupational exposure incident see Appendix 10.

Safety devices

Health and Safety (Sharp Instruments in Healthcare) Regulations 2013 are concerned with reducing and eliminating the number of 'sharps' related injuries which occur within healthcare. Its basic guidance is:

- avoid unnecessary use of sharps;
- if use of medical sharps cannot be avoided, source and use a 'safer sharp' device:
- if a safer sharp device is not available then safe procedures for working with and disposal must be in place eg sticky mats, sharps bins, safety procedures and training.

Sharps handling must be assessed, kept to a minimum and eliminated, if possible, with the use of approved safety devices.

- Manufacturers' instructions for safe use and disposal must be followed.
- Needles must not be re-sheathed/recapped or disassembled after use.
- Sharps must not be passed directly hand to hand.
- Used sharps must be discarded at the point of use by the person generating the waste.
- Always dispose of needles and syringes as 1 unit.
- If a safety device is being used safety mechanisms must be deployed before disposal.

When transporting sharps boxes for community use these must be transported safely with the use of temporary closures.

Further information can be found in <u>occupational exposure management (incl. sharps)</u> literature review.

Chapter 2: Transmission based precautions (TBPs)

SICPs may be insufficient to prevent cross transmission of specific infectious agents and additional precautions called "Transmission Based Precautions" (TBP) may be required when caring for patients with known / suspected infection or colonisation.

Transmission Based Precautions are categorised by the route of transmission of infectious agents (some infectious agents can be transmitted by more than one route).

Clinical judgement and decisions should be made by staff on what additional precautions are required and this will be based on:

- suspected/known infectious agent
- severity of the illness caused
- transmission route of the infectious agent
- care setting and procedures undertaken.

Type of precautions:

Contact precautions:

Used to prevent and control infections that spread via direct contact with the patient or indirectly from the patient's immediate care environment (including care equipment). This is the most common route of cross-infection transmission.

Droplet precautions:

Measures used to prevent, and control infections spread over short distances (at least 1 meter)3 via droplets from the respiratory tract of one individual directly onto a mucosal surface or conjunctivae of another individual. Droplets penetrate the respiratory system to above the alveolar level.

3. During the COVID-19 pandemic increased physical distancing (2 metres) was introduced as an additional IPC measure. This has now decreased to pre-pandemic physical distancing (1 metre) in all areas.

Airborne precautions:

Measures used to prevent, and control infection spread without necessarily having close patient contact via aerosols from the respiratory tract of one individual directly onto a mucosal surface or conjunctivae of another individual. Aerosols can penetrate the respiratory system to the alveolar level.

The traditional modes of transmission for respiratory infectious agents as defined before the COVID-19 pandemic are unlikely to be as delineated as is described in the scientific literature, i.e., droplet or airborne transmission and the application of TBPs may differ

depending on the setting and the known or suspected infectious agent. Applications of TBPs should be considered within the framework of the hierarchy of controls. Settingspecific risk assessment tools are available to support organisations in applying the HoC https://www.england.nhs.uk/publication/national-infection-prevention-and-control/.

Appendix 11 provides details of the type of precautions, optimal patient placement, isolation requirement and respiratory precautions required.

Further information on Transmission Based Precautions can be found in the definitions of Transmission Based Precautions literature reviews.

NB During the COVID-19 pandemic universal masking in healthcare and increased physical distancing were introduced as additional IPC measures. These measures are not included in this version of the NIPCM as it is currently covered by UK IPC Guidance as published by UKHSA.

2.1 Patient placement / assessment of infection risk

The potential for transmission of infection must be assessed when a patient enters a care area. If hospitalised/in a care home setting, this should be continuously reviewed throughout the stay/period of care. The assessment should influence patient placement decisions in line with clinical/care need(s).

Patients who may present a cross-infection risk in any setting includes those:

- with diarrhoea, vomiting, an unexplained rash, fever or respiratory symptoms
- known to have been previously positive with multidrug-resistant organisms (MDRO) eg MRSA, CPE
- who have been an inpatient in any hospital in the UK or abroad or are a known epidemiological link to a carrier of CPE
- who have a known or suspected infection or colonisation.

Isolation facilities should be prioritised depending on the known/suspected infectious agent (refer to the aides memoire in this document – appendices 11a and 11b).

All patient placement decisions and assessment of infection risk (including isolation requirements) must be clearly documented in the patient notes and provided in patient handovers with other healthcare/ care providers.

The clinical judgement and expertise of the staff involved in a patient's management and the Infection Prevention and Control Team (IPCT) should be sought, particularly for the application of TBPs, eg isolation prioritization, when single rooms are in short supply.

Single room isolation in hospital settings:

- Isolation of infectious patients can be in specialised isolation facilities, single room isolation, cohorting of infectious patients where appropriate, ensuring that they are separated by at least 3 feet (1 metre) with the door closed.
- Signage should be used on doors/areas to communicate isolation requirements and prevent entry of unnecessary visitors, non-essential staff. Patient confidentiality must be maintained.
- Infectious patients should only be transferred to other departments if clinically necessary. If the patient has an infectious agent transmitted by the airborne/droplet route, then if possible/tolerated the patient should wear a surgical face mask in communal areas during transfer.
- Receiving department/hospital and transporting staff must be aware of the necessary precautions.
- Isolation room doors should remain closed, if this is not possible, eg paediatrics, there should be a documented risk assessment.
- If single rooms are limited, infectious patients who have conditions that could increase the risk of transmission of infection to other patients, such as, excessive cough or an MDRO should be prioritised for placement in a single room.
- Single room prioritisation should be reviewed daily and the clinical judgement and expertise of the staff involved in a patient's management and the Infection Prevention and Control team or Health Protection Team should be sought particularly for the application of TBPs

Cohorting in hospital settings:

Cohorting of infectious patients can be considered when:

- Single rooms are in short supply and there are two or more patients (a cohort) with the same confirmed infection
- There are situations of service pressure e.g. winter and patients may have different or multiple infections. In these situations, a preparedness plan must be in place ensuring the organisational/Board level assurance on IPC systems and processes are in place (e.g. IPC BAF)

Infectious patients must not be cohorted with others with different of multiple infections include:

- Those at increased risk of acquisition and adverse outcomes resulting from infection (e.g. immunosuppression)
- Individuals who are unlikely to comply with TBPs

Single room isolation in care settings with or without nursing care:

- Residents should remain in their bedroom while considered infectious and the door should remain closed (if unable to isolate this should be documented).
- If transfer to hospital is required, ambulance services and the hospital admission area should be informed of the infectious status of the resident. Advice on resident's clinical management should be sought from the GP and the local

health protection unit infection prevention team.

Avoid unnecessary transfer of residents within/between care areas.

Primary care/outpatient settings:

Patients attending with suspected/known infection/colonisation should be prioritised for assessment/treatment, eg scheduled appointments at the start or end of the clinic session.

Infectious patients should be separated from other patients while awaiting assessment and during care management by at least 3 feet (1m).

If transfer from a primary care facility to hospital is required, ambulance services should be informed of the infectious status of the patient. Patient confidentiality must be maintained.

Staff cohorting: consider assigning a dedicated team of care staff to care for patients in isolation/cohort rooms/areas as an additional infection control measure during outbreaks/incidents. This can only be implemented if there are sufficient levels of staff available (so as not to have a negative impact on non-affected patients' care).

Before discontinuing isolation:

Individual patient risk factors should be considered (eg there may be prolonged shedding of certain microorganisms in immunocompromised patients).

2.2 Safe management of patient care equipment in an isolation room/cohort area

- Use single-use items if possible.
- Reusable non-invasive care equipment should be dedicated to the isolation room/cohort area and decontaminated prior to use on another patient.
- An increased frequency of decontamination should be considered for reusable non-invasive care equipment when used in isolation/cohort areas.
- For how to decontaminate non-invasive reusable equipment see Appendix 7.

2.3 Safe management of the care environment

The care environment must be:

- visibly clean, free from non-essential items and equipment to facilitate effective cleaning
- well maintained, in a good state of repair and with adequate ventilation for the clinical specialty.

Equipment used for environmental decontamination must be either single-use or dedicated to the affected area then decontaminated or disposed of following use eg cloths, mop heads.

Environmental decontamination: enhanced cleaning

Refer to the National Cleaning Standards for enhanced cleaning in different settings.

Inpatient settings:

Patient isolation/cohort rooms/area must be decontaminated at least daily, this may be increased on the advice of IPCTs/. These areas must be decontaminated using either:

- a combined detergent/disinfectant solution at a dilution of 1,000 parts per million available chlorine (ppm available chlorine (av.cl.)); or
- a general-purpose neutral detergent in warm water followed by solution of 1,000ppm av.cl.

Alternative cleaning agents/disinfectant products may be used with agreement of the local IPC team.

Employers must ensure that cleaning products and protocols are managed and risk assessed in accordance with the COSHH regulations - Control of substances hazardous to health (COSHH) - health and safety topics in cleaning (hse.gov.uk).

Manufacturers' quidance and recommended product 'contact time' must be followed for all cleaning/disinfection solutions.

Increased frequency of decontamination/cleaning schedules should be incorporated into the environmental decontamination schedules for areas where there may be higher environmental contamination rates, eg:

- toilets/commodes particularly if patients have diarrhoea; and
- "frequently touched" surfaces eg door/toilet handles, locker tops, over bed tables and bed rails.

Vacated rooms should also be decontaminated following an AGP. Clearance of infectious particles after an AGP is dependent on the ventilation and air change within the room. This is a minimum of 20 minutes in hospital settings where the majority of these procedures occur. In general wards and single rooms there should be a minimum of 6 air changes per hour, in negative-pressure isolation rooms there should be a minimum of 10 air changes per hour. Advice should be sought from IPCT.

Primary care/outpatient settings:

The extent of decontamination between patients will depend on the duration of the consultation/assessment, the patients presenting symptoms and any visible environmental contamination.

Terminal decontamination

Following patient transfer, discharge, or once the patient is no longer considered infectious, remove from the vacated isolation room/cohort area, all:

- healthcare waste and any other disposable items (bagged before removal from the room)
- bedding/bed screens/curtains manage as infectious linen (bagged before removal from the room)
- reusable non-invasive care equipment (decontaminated in the room prior to removal) Appendix 7.

The room should be decontaminated using either:

- a combined detergent disinfectant solution at a dilution (1,000ppm av.cl.); or
- a general-purpose neutral detergent in warm water followed by a solution of 1,000ppm av.cl. (or alternative locally agreed cleaning product)

Rooms must be cleaned from highest to lowest points and from least to most contaminated points.

Organisations can consider using Hydrogen Peroxide Vapour disinfection or ultra-violet light technology for specific pathogens. Manufacturers' guidance and recommended product "contact time" must be followed for all cleaning/disinfection solutions.

Terminal cleaning of outpatients/theatre recovery areas should be in accordance with local policy advised by the IPCT.

2.4 Personal protective equipment (PPE): respiratory protective equipment (RPE)

Personal Protective Equipment (PPE) must still be used in accordance with SICPs when using Respiratory Protective Equipment (RPE). See Chapter 1.4 for PPE use for SICPs.

Where it is not reasonably practicable to prevent exposure to a substance hazardous to health (as may be the case where healthcare workers are caring for patients with suspected or known airborne pathogens), the hazard must be adequately controlled by applying protection measures appropriate to the activity and consistent with the assessment of risk.

If the hazard is unknown the clinical judgement and expertise of IPC staff is crucial and the precautionary principle should apply.

Respiratory Protective Equipment (RPE) i.e., a filtering face piece (FFP) must be considered when a patient is admitted with a known/suspected infectious agent/disease spread wholly or partly by the airborne route and when carrying out aerosol generating procedures (AGPs) on patients with a known/suspected infectious agent spread wholly or partly by the airborne or droplet route.

Staff in primary care/outpatient settings or care homes would not normally be required to wear an FFP3 respirator for routine care unless an AGP is being performed on an infectious patient in which case staff should wear a fit tested, FFP3 respirator.

The decision to wear an FFP3 respirator/hood should be based on clinical risk assessment eg task being undertaken, the presenting symptoms, the infectious state of the patient, risk of acquisition and the availability of treatment.

For a list of organisms spread wholly or partly by the airborne (aerosol) or droplet routes see Appendix 11a.

National Priority Risk Categorisation for fit testing with FFP3 respirators

The following risk categorisation is the minimum requirement for staff groups that require FFP3 respirator fit testing. Healthcare organisations can add to this, for

example where there are high risk units. This categorisation is inclusive of out of hours services

Level 1 – Preparedness for business as usual

Staff in clinical areas most likely to provide care to patients who present at healthcare facilities with an infectious pathogen spread by the airborne route; and/or undertake aerosol generating procedures i.e., A&E, ICU, paediatrics, respiratory, infectious diseases, anaesthesia, theatres, chest physiotherapists, Special Operations Response Team (Ambulance), A&E, ambulance staff, bronchoscopy staff, resuscitation teams, mortuary staff.

Level 2 – Preparedness in the event of emerging threat

Staff in clinical settings likely to provide care to patients admitted to hospital in the event of an emerging threat eg Medical receiving, Surgical, Midwifery and Specialty wards, all ambulance staff. In the event of an 'Epidemic/Pandemic' Local Assessment as per organizations preparedness plans apply.

FFP3 respirator or powered respirator hood:

- may be considered for use by visitors if there has been no previous exposure to the infected person or infectious agent; but
- must never be worn by an infectious patient(s) due to the nature of the respirator filtration of incoming air not expelled air.
- powered respirator hoods are an alternative to tight-fitting FFP3 respirators for example when fit testing cannot be achieved.
- powered hoods can be single use (disposable) or reusable (with a decontamination schedule, see note) and must be fluid-resistant; the filter must be enclosed with the exterior and the belt able to withstand disinfection with 10,000ppm av.cl.
- Respirators and powered respirator hoods with exhalation valves are ineffective for source control. These should not be worn by a healthcare worker/operator when sterility directly over the surgical field is required eg in theatres/surgical settings or when undertaking a sterile procedure (see National Patient Safety Alert).

All tight-fitting RPE i.e., FFP3 respirators must be:

- single-use (disposable) or reusable, and preferably fluid resistant. (if not a full face visor should be worn)
- fit tested on all healthcare staff who may be required to wear a respirator to ensure an adequate seal/fit according to the manufacturers' guidance.
- fit checked (according to the manufacturers' guidance) every time a respirator is donned to ensure an adequate seal has been achieved.
- compatible with other facial protection used i.e. protective eyewear so that this does not interfere with the seal of the respiratory protection. Regular corrective spectacles are not considered adequate eye protection.

HSE guidance and demonstrations for putting on respirators and performing a fit check is available here.

For any facial hair, the hair must not cross or interfere with the respirator sealing surface. If the respirator has an exhalation valve, hair within the sealed mask area should not

impinge upon or contact the valve. You must pass a face fit test for any tight-fitting respiratory protective equipment that you need to use for work activities.

Please note: reusable respirators including powered respirator hoods must comply with HSE guidance (HSG53) and be adequate and suitable for their intended use. Reusable respirators must have a decontamination schedule in place and be maintained according to manufacturer's instructions.

Further information regarding fitting and fit checking of respirators can be found on the Health and Safety Executive website at: <a href="http://www.hse.gov.uk/respiratory-protective-prote equipment/basics.htm

Removal (doffing) of PPE

- In the absence of an anteroom/lobby remove FFP3 respirators and eye/face protection in a safe area (eg outside the isolation/cohort room/area).
- All other PPE should be removed in the patient care area.

For the recommended method of putting on and removing PPE, see <u>UKHSA guides</u>.

Further information can be found in Respiratory Protective Equipment (RPE) literature review and PPE for Infectious Diseases of High Consequence (IDHC) literature review.

Aerosol generating procedures

Aerosol generating procedures (AGPs) are medical procedures that can result in the release of aerosols from the respiratory tract. The criteria for an AGP are a high risk of aerosol generation and increased risk of transmission (from patients with a known or suspected respiratory infection).

The list of medical procedures that are considered to be aerosol generating and associated with an increased risk of respiratory transmission is:

- awake* bronchoscopy (including awake tracheal intubation)
- awake* ear, nose, and throat (ENT) airway procedures that involve respiratory suctioning
- awake* upper gastro-intestinal endoscopy
- dental procedures (using high speed or high frequency devices, for example ultrasonic scalers/high speed drills)
- induction of sputum
- respiratory tract suctioning**
- surgery or post-mortem procedures (like high speed cutting / drilling) likely to produce aerosol from the respiratory tract (upper or lower) or sinuses.
- tracheostomy procedures (insertion or removal).

- *Awake including 'conscious' sedation (excluding anesthetized patients with secured airway)
- ** The available evidence relating to respiratory tract suctioning is associated with ventilation. In line with a precautionary approach, open suctioning of the respiratory tract regardless of association with ventilation has been incorporated into the current (COVID-19) AGP list. It is the consensus view of the UK IPC cell that only open suctioning beyond the oro-pharynx is currently considered an AGP, that is oral/pharyngeal suctioning is not an AGP.

Further information can be found in the rapid review of aerosol generating procedures.

Infection prevention and control when caring for the deceased

The principles of SICPs and TBPs continue to apply while deceased individuals remain in the care environment. This is due to the ongoing risk of infectious transmission via contact although the risk is usually lower than for living patients.

Staff should advise relatives of the precautions following viewing and/or physical contact with the deceased and also when this should be avoided.

Washing and/or dressing of the deceased should be avoided if the deceased is known or suspected to have an invasive streptococcal infection, viral hemorrhagic fevers or other Hazard Group 4 infectious agents. See Appendix 11b.

Deceased patients with a suspected or confirmed Hazard Group 4 pathogen should be transferred to a specialised high consequence infectious disease (HCID) facility as part of the HCID response network.

Deceased individuals known or suspected to have a Hazard Group 4 infectious agent should be placed in a sealed double plastic body bag with absorbent material placed between each bag. The surface of the outer bag should be disinfected with 1000ppm av.cl before being placed in a robust sealed coffin.

Post-mortem examination should not be performed on a deceased individual known or suspected to have Hazard Group 4 infectious agents. See Appendix 11b. Blood sampling can be undertaken in the mortuary by a competent person to confirm or exclude this diagnosis. Refer to Section 2.4 for suitable PPE.

Further information can be found in the infection prevention and control during care of the deceased literature review

Refer to HSG283 – managing infectious risk when handling the deceased for more information.

Appendix 1: Best practice - How to hand wash, step-by-step images



Best Practice: How to hand wash step by step images

Steps 3-8 should take at least 15 seconds.



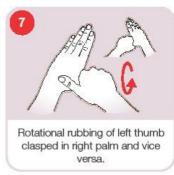














hand in left palm and vice versa.









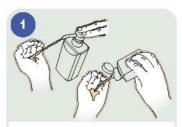
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*Any skin complaints should be referred to local occupational health or GP.

Appendix 2: Best practice – How to handrub, step-by-step images



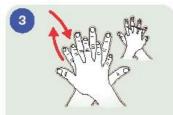
Best Practice: How to handrub step by step images



Apply a palmful of the product in a cupped hand and cover all surfaces.



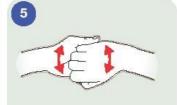
Rub hands palm to palm.



Right palm over the back of the other hand with interlaced fingers and vice versa.



Palm to palm with fingers interlaced.



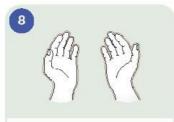
Backs of fingers to opposing palms with fingers interlocked.



Rotational rubbing of left thumb clasped in right palm and vice versa.



Rotational rubbing, backwards and forwards with clasped fingers of right hand in left palm and vice versa.



Once dry, your hands are safe.

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Appendix 3: Best practice - surgical hand antisepsis using antimicrobial soap





Put antimicrobial liquid soap onto the palm of each hand/arm using the elbow of your other arm to operate the dispenser.



Rub hands palm to palm. Steps 3-8 should take a minimum of 2 minutes.



Right palm over the back of the other hand with interlaced fingers and vice versa.



Palm to palm with fingers interlaced.



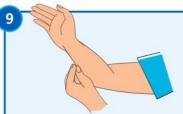
Backs of fingers to opposing palms with fingers interlaced.



Rotational rubbing of left thumb clasped in right palm and vice versa.



Rotational rubbing, backwards and forwards with clasped fingers of right hand in left palm and vice versa. Rinse hands between steps 8-9, passing them through the water in one direction only.



Put antimicrobial liquid soap onto the palm of your left hand using elbow of your other arm to operate the dispenser. Use this to scrub the right arm for 1 minute using a rotational method keeping the hand higher than the arm at all times.

Repeat the process for the other hand and arm keeping hands above elbows at all times.

If the hand touches anything at any time, the scrub must be lengthened by 1 minute for the area that has been contaminated.

*Nails should be cleaned using a soft, single-use disposable nail brush or a nail pick before the first scrub of the day or if visibly dirty. Any skin complaints should be referred to local occupational health or GP.

> Local policy may recommend repeating steps 1-11 to the mid-forearms only.



Rinse hands and arms by passing them through the water in one direction only, from fingertips to elbow. Do not move the arm back and forth through the water.

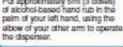


Hold hands above the elbow. Use one sterile, disposable towel per hand and arm. Blot the skin of the hand, then use a corkscrew movement to dry from the hand to the elbow. The towel must not be returned to the hand once the arm has been dried and must be discarded immediately.

Appendix 4: Best practice - surgical hand rub technique using alcohol based hand rub (ABHR)

- The hand rubbing technique for surgical hand preparation must be performed on clean, dry hands.
- On arrival in the operating theatre and after having donned theatre clothing (cap/hat/bonnet and mask). hands must be washed with soap and water.
- After the operation when removing gloves, hands must be rubbed with an alcohol-based formulation or washed with soap and water if any residual talc or biological fluids are present (e.g. the glove is punctured).
- · Surgical procedures may be carried out one after the other without the need for hand washing, provided that the hand rubbing technique for surgical hand preparation is followed (images 1 to 15).







Dip the fingertips of your right hand in the hand rub to decontaminate under the nails



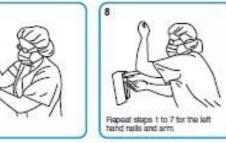




skin area is covered by using circular movements around the forearm until the hand rub has fully evaporated (10-15 seconds).









Put approximately Smi (3 doses) of alcohol-based handrub in the pain of your left hand, using the elbow of your other arm to operate the distributor. Rub both hands in the same time up to the wrists, and ensure that all the sleps presented in steps 9-14 are followed.



Cover the whole surface of the hands up to the wrist with alcohol-based hand rub, rubbing palm against palm with a rotatin

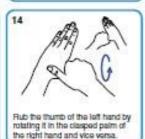


Rub the back of the hands up to the wrist with alcohol-based handrub, rubbing palm against palm with a rotating movement



Rub the back of the left hand, including the wrist, moving the n back and forth and











Appendix 5a: Personal protective equipment (PPE) when applying standard infection control precautions (SICPs)

Before undertaking any procedure or task, staff should assess any likely exposure to blood and/or other body fluids, non-intact skin, mucous membranes or any equipment or items in the care environment that could be contaminated and wear personal protective equipment (PPE) if required. PPE must protect adequately against the risks associated with the procedure or task.

Hand hygiene must be performed before putting on and after removal of PPE.

| SICPs | Gloves | Apron | Gown (ambulance staff use coveralls) | Fluid resistant surgical mask (FRSM) | Eye/face protection |
|--|--------|-------|--|--|------------------------|
| No anticipated exposure to blood or body fluid, mucous membranes, or non-intact skin. | 8 | 8 | 8 | 8 | 8 |
| Exposure to blood or body fluid, mucous membranes, or non-intact skin is anticipated but NO risk of splashing or spraying. | | | 8 | 8 | 8 |
| Exposure to blood or body fluid, mucous membranes, or non-intact skin is anticipated AND risk of spraying or splashing. | | | Unless in place of an apron if extensive spraying or splashing is anticipated. | | |

Where to put on and remove PPE

If required as above, PPE should be put on within the patient room/care area.

Gloves are not an alternative to hand hygiene. Gloves must always be removed after each task on the same patient and hand hygiene performed as per the 5 moments for hand hygiene.

All PPE must be removed and disposed of before leaving the patient room/care area on completion of care episode.

NB. Universal masking using FRSM may be indicated as a source control measure during outbreaks of respiratory infectious agents.

Appendix 5b: Personal protective equipment (PPE) when applying transmission based precautions (TBPs)

SICPs may be insufficient to prevent cross transmission of specific infectious agents and additional precautions (TBPs) may be required. PPE must protect adequately against the risks associated with the procedure or task. Refer to appendix 11a for additional information.

Hand hygiene must be performed before putting on and after removal of PPE.

| TBPs | Gloves | Apron | Gown | Fluid resistant surgical mask (FRSM) | Respiratory Protective Equipment (RPE) | Eye/face protection |
|------------------------|--|----------|---|--|--|--|
| Contact precautions | Unless exposure to blood or body fluid, mucous membranes, or non-intact skin is anticipated or footnote 1 applies¹ | | Unless in place of an apron if extensive spraying or splashing is anticipated | Unless risk of splashing or spraying of blood or body fluids is anticipated or footnote 2 applies ² | 8 | Unless risk of splashing or spraying of blood or body fluids is anticipated |
| Droplet precautions | | Ø | Unless in place of an apron if extensive spraying or splashing is anticipated | | 8 | |
| Airborne precautions | Ø | 8 | Ø | 8 | Ø | Ø |

Where to put on and remove PPE

Gloves are not an alternative to hand hygiene. Gloves must always be removed after each task on the same patient and hand hygiene performed as per the 5 moments for hand hygiene.

Contact precautions: required PPE should be put on within the patient room/care area immediately before direct contact with the patient or their environment and should be removed and disposed of before leaving the patient room/care area.

Droplet and airborne precautions: required PPE should be put on before entering the patient room/care area. Unless there is a dedicated isolation room with anteroom, gowns, aprons and gloves should be removed and disposed of before leaving the patient room/care area. Eye/face protection and RPE (if worn) must be removed and disposed of after leaving the patient room/care area.

PPE requirements for high consequence infectious diseases should be discussed with specialist teams as per appendix 11b.

^{1.}Clinical risk assessment may also indicate the use of gloves for specific organisms such as scabies, multi-drug resistant organisms or those with increased potential for hand and environmental contamination such as spore forming organisms e.g. *C. difficile*. This list is not exhaustive.

2.Universal masking using FRSM may be indicated as a source control measure during outbreaks of respiratory

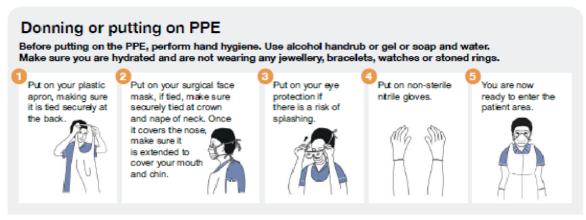
infectious agents.

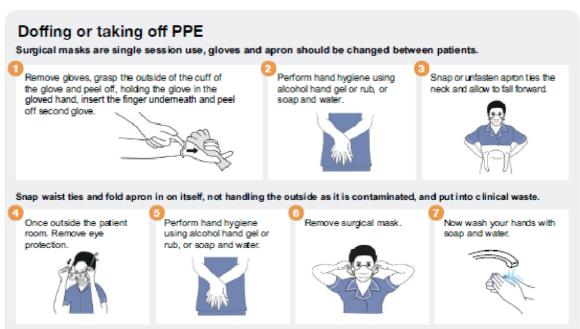
Appendix 6: Putting on and removing PPE (donning and doffing)



Guide to donning and doffing PPE: **Droplet Precautions**

for health and social care settings





Please refer to the PHE standard PPE video in the COVID-19 guidance collection: www.gov.uk/government/publications/covid-19-personal-protective-equipment-use-for-non-aerosol-generating-procedures

Appendix 7: Best practice – decontamination of reusable non-invasive care equipment

| Infection prevention team/health protecti | | Routine decontamination of reusable non-invasive care equipment. | | | |
|--|--|--|--|--|--|
| Name: | | | | | |
| Designation: Contact Number: | | Check manufacturer's instructions for suitability of cleaning products especially when dealing with electronic equipment. Wear appropriate PPE eg disposable, non-sterile gloves and apron. | | | |
| | | , | <u> </u> | | |
| Is equipment contaminated vomit/faeces or been used a known/suspected infection | on a patient with | ← NO | Is equipment contaminated with blood or body fluid as in box 1? | | |
| NO 🗸 | YES | | YES | | |
| Decontaminate equipment with disposable cloths/paper towel and a fresh solution of general purpose detergent and water or detergent impregnated wipes. Rinse and dry thoroughly. Disinfect specific items of non-invasive, reusable, communal care equipment if recommended by the manufacturer's instructions eg stethoscopes with 70% isopropyl alcohol. | Immediately deception equipment with cloths/paper tow solution of deter and follow with solution of 1,000 million availab (ppm av cl)* rinthoroughly. Or use a combin /chlorine releasin with a concentra ppm av cl* rinst thoroughly. *If the item cannochlorine releasing with the manufact instructions for a salternative to use foombined with deceaning. | disposable vel and a fresh rgent, rinse dry a disinfectant D parts per le chlorine use and dry ed detergent ug solution ation of 1,000 e and dry t withstand agents, consult turer's uitable following or | Immediately decontaminate equipment with disposable cloths/paper towel and a fresh solution of detergent, rinse dry and follow with a disinfectant solution of 10,000 parts per million available chlorine (ppm av cl)*. Or use a combined detergent /chlorine releasing solution with a concentration of 10,000 ppm av cl* rinse and dry thoroughly. *If the item cannot withstand chlorine releasing agents, consult with the manufacturer's instructions for a suitable alternative to use following or combined with detergent cleaning. | | |
| $\overline{}$ | $\overline{}$ | | | | |
| ☐ Follow manufacturer's instructions for safe preparation, dilution, and contact time. ☐ Clean the piece of equipment from the top or furthest away point. ☐ Discard disposable cloths/paper roll immediately into the healthcare waste receptacle. ☐ Discard detergent/disinfectant solution in the designated area. ☐ Clean, dry and store re-usable decontamination equipment. ☐ Remove and discard PPE. ☐ Perform hand hygiene. BOX 1 • Cerebrospinal fluid • Synovial fluid • Vaginal secretions | | | | | |
| Peritoneal fluid Pleural fluid | Amniotic fluid | Breast milk | s fluid with visible blood (excluding urine) | | |

Appendix 8: Best practice – linen bagging and tagging

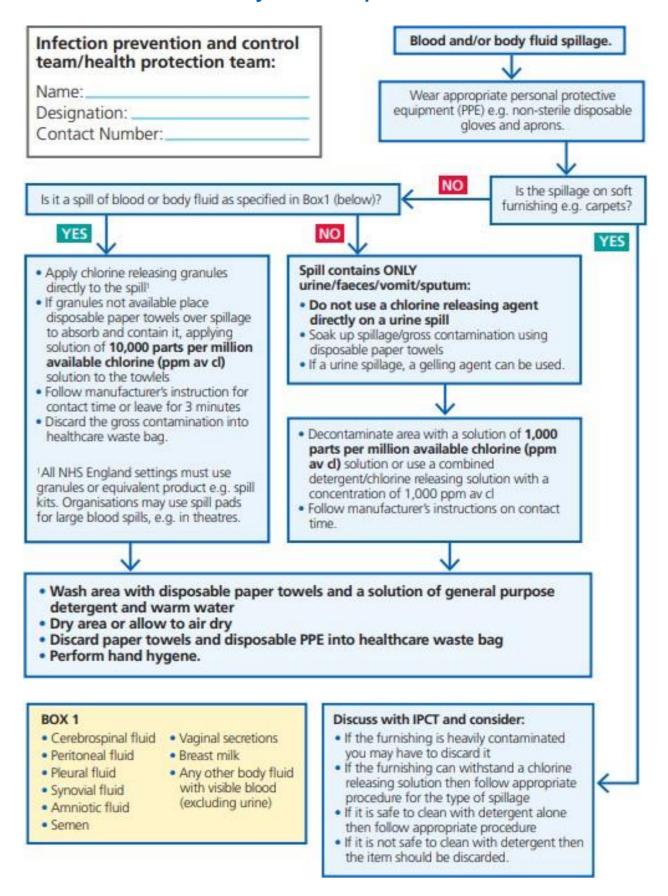
CATEGORY INNER BAG LINEN BAG/HAMPER **Used linen** None required (All used linen in the care setting not contaminated by blood or body fluids) Water soluble or Infectious linen soluble seam (All linen used by a person known or (alginate bag) suspected to be placed into a polythene infectious and linen that is bag contaminated with blood or body fluids) If 'used' none Heat-labile linen required. Linen that may be damaged If 'infectious' treat (shrinkage/ stretching) by as above. COLOUR thermal MAY disinfection.

Ensure all linen bags are:

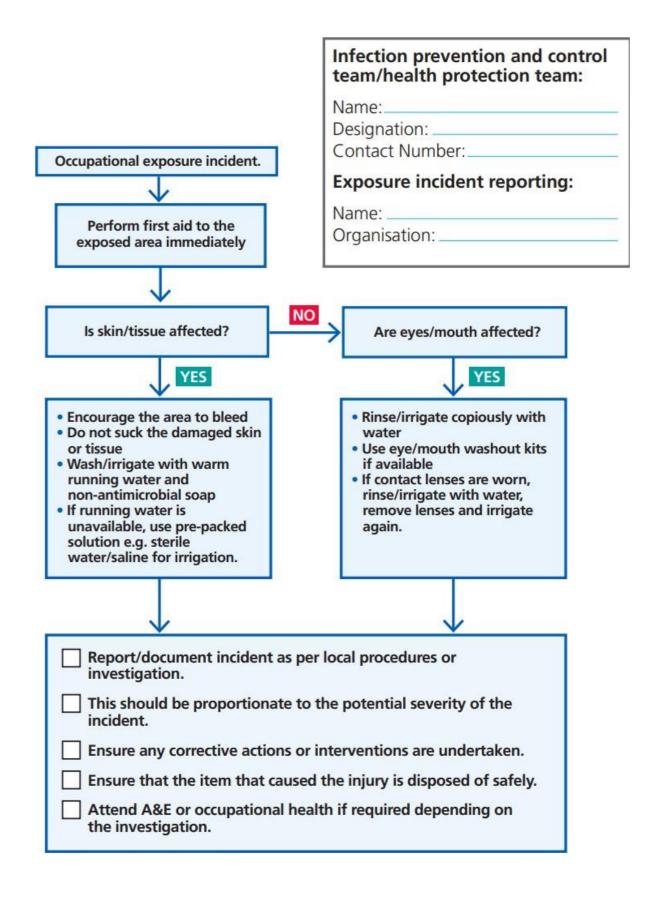
- Securely tied
- Not overfilled
- Tagged with hospital, ward/department and date.

If linen processing has been outsourced to an industrial service provider, follow the categorisation system and colour coding for 'used' and 'infectious' linen set out by the service provider.

Appendix 9: Best practice – management of blood and body fluid spills



Appendix 10: Best practice - management of occupational exposure incidents



Appendix 11a: Aide memoire for optimal patient placement and respiratory protective equipment (RPE) for infectious agents in hospital inpatients (based on evidence from WHO, CDC and UKHSA)

The clinical judgement and expertise of the IPC and Health Protection Teams should be sought for novel, unusual pathogens or where an increase in cases has been detected in any care setting. Advice can also be sought from the bacterial reference departments at UKHSA for rare / unusual pathogens, exceptional phenotypes or for advice regarding typing of outbreak strains.

The following table outlines the transmission-based precautions (TBPs) required for several infectious agents / diseases which will minimise cross transmission events from and between patients, and healthcare workers. The details included in the table below are drawn from published evidence from a number of validated sources, for example, WHO, CDC, and UKHSA. The table is intended to function as a quick reference quide, is not exhaustive, and is not intended to replace appropriate risk assessment and clinical judgement or formal assessments by public health agencies. The table summarises:

- Optimal patient placement while the patient is considered infectious; and
- The recommended RPE (recognising other PPE is required) to minimise risk of cross infection to staff, patients and visitors.
- Decisions made by staff regarding use/non-use of RPE will depend on the completion of clinical risk assessment. considering presenting symptoms, available treatments, the risk of acquisition, the level of interaction, task to be performed, and / or the anticipated level of exposure to blood and / or other body fluids.
- In the hospital setting patients with suspected or confirmed respiratory symptoms should, whenever possible, be placed in a single room, ideally with en-suite facilities If a single / isolation room is not available, cohort patients with confirmed respiratory infection with other patients confirmed to have the same infectious agent. Patients with suspected or confirmed respiratory infection should be provided with a surgical facemask (Type II or Type IIR) to be worn in multi- bedded bays and communal areas if this can be tolerated, and where the patient cannot be isolated in a single room.

• Note: *The distinction between droplet and aerosol transmission is not always clearly defined. A dynamic clinical risk assessment should be performed using the hierarchy of controls to inform the assessment and should include evaluation of the ventilation in the area, operational capacity, and prevalence of infection in the local area. Staff should be provided with training on the correct use of RPE. Current guidance is that an FFP3 respirator must be worn by staff when caring for patients with a suspected or confirmed infection spread by the airborne route, when performing AGPs on a patient with a suspected or confirmed infection spread by the droplet or airborne route, and when deemed necessary after risk assessment.

| Suspected or confirmed Pathogen | Disease | Transmission based precautions (TBPs) required | Optimal placement while patient is considered infectious and until resolution of symptoms | Respiratory protection (RPE) for healthcare workers while patient is considered infectious ³ | Notifiable under Public Health Act 1984 and Health Protection Regulations 2010 ⁴ |
|---|---|---|--|--|---|
| Acinetobacter baumannii | Pneumonia, bacteraemia, skin and soft tissue infections. | Contact | Single en-suite room in high risk settings eg ICU/PICU/NICU, oncology/haematology | No requirement for RPE | No |
| Acute infectious hepatitis of unknown aetiology | Acute hepatitis | Droplet | Single en-suite room | Fluid resistant surgical facemask (FRSM) for routine care and FFP3 or Hood for AGPs | Yes |
| Adenovirus ¹ | Upper +/- lower respiratory tract infection | Droplet | Single en-suite room | Fluid resistant surgical facemask (FRSM) for routine care and FFP3 or Hood for AGPs | No |
| | Conjunctivitis, gastroenteritis | Contact | Single en-suite room | No requirement for RPE | No |
| Bacillus anthracis | Respiratory, gastrointestinal or cutaneous Anthrax | Contact - Enteric if the patient has diarrhoea | Single en-suite room | No requirement for RPE ⁶ | Yes |

| Suspected or confirmed Pathogen | Disease | Transmission based precautions (TBPs) required | Optimal placement while patient is considered infectious and until resolution of symptoms Respiratory protection (RPE) for healthcare workers while patient is considered infectious ³ | | Notifiable under Public Health Act 1984 and Health Protection Regulations 2010 ⁴ |
|---|--|---|--|---|---|
| Bacillus cereus | Gastroenteritis, sepsis, pneumonia, endocarditis, central nervous system (CNS) and ocular infections | Contact Enteric if the patient has diarrhoea | Single en-suite room in high risk settings eg ICU/PICU/NICU, oncology/haematology | igh risk settings eg ICU/PICU/NICU, No requirement for RPE | |
| Bordetella pertussis | Whooping Cough | Droplet | Single en-suite room | Fluid resistant surgical facemask (FRSM) for routine care and FFP3 or Hood for AGPs until patient has been established on appropriate antimicrobial treatment ⁵ | Yes |
| Candida auris | Ear, wound and bloodstream infection | Contact | Single en-suite room in high risk settings eg ICU/PICU/NICU, oncology/haematology | No requirement for RPE | No |
| Carbapenemase producing Enterobacterales (CPE) (either swab positive or positive as per clinical risk assessment criteria | Colonisation, device associated infections – urinary tract infection, catheter associated bacteraemia | Contact | Single en-suite room | No requirement for RPE | No |
| Chlamydia pneumoniae | Pneumonia | Droplet | Single en-suite room in high risk settings eg ICU/PICU/NICU, oncology/haematology | Fluid resistant surgical facemask (FRSM) for routine care and FFP3 or Hood for AGPs | No |
| Clostridioides difficile | Clostridioides difficile infection (CDI) | Enteric | Single en-suite room | No requirement for RPE | No |
| Coronavirus ¹ (Seasonal) including SARS- CoV-2 | Respiratory symptoms including asymptomatic presentations COVID-19 | Droplet/Airborne * please see note above | Single en-suite room | Fluid resistant surgical facemask (FRSM) for routine care and FFP3 or Hood for AGPs* please see note above | No Yes SARS- CoV-2 |

| Suspected or confirmed Pathogen | Disease | Transmission based precautions (TBPs) required | Optimal placement while patient is considered infectious and until resolution of symptoms | Respiratory protection (RPE) for healthcare workers while patient is considered infectious ³ | Notifiable under Public Health Act 1984 and Health Protection Regulations 2010 ⁴ |
|---|---|---|--|--|---|
| Corynebacterium diphtheria or Corynebacterium ulcerans | Diphtheria – Cutaneous, Pharyngeal (toxigenic strains) | Contact, Droplet (If Pharyngeal) | Single en-suite room | Fluid resistant surgical facemask (FRSM) for routine care and FFP3 or Hood for AGPs (if pharyngeal) | Yes |
| Enterovirus D68 | Mild to moderate upper respiratory tract infections. Can cause severe respiratory illness and rarely acute flaccid myelitis (AFM) | Droplet | t Single en-suite room Fluid resistant surgical facemask (FRSM) for routine care and FFP3 or Hood for AGPs | | No |
| Gastrointestinal infections eg Salmonella spp. | Gastroenteritis | Enteric | Single en-suite room | Fluid resistant surgical facemask (FRSM) if vomiting is present. | (Some GI Infections are notifiable. Refer to guidance) |
| Haemophilus influenzae (all invasive*) | Epiglottitis, *meningitis, pneumonia, *bacteraemia | Droplet | Single en-suite room | Fluid resistant surgical facemask (FRSM) for routine care and FFP3 or Hood for AGPs until patient has been established on appropriate antimicrobial treatment ⁵ | Yes *Only |
| Hepatitis A virus | Hepatitis, Gastroenteritis | Enteric | Single en-suite room | Fluid resistant surgical facemask (FRSM) if vomiting is present. | Yes |
| Herpes zoster (Shingles) (varicella-zoster) ² | Shingles (vesicle fluid) | Contact | Single en-suite room If lesions cannot be covered | No requirement for RPE | Notifiable organism but not notifiable disease |
| | Disseminated zoster | Airborne | Isolation room/suite | Fluid resistant surgical facemask (FRSM) for routine care and FFP3 or Hood for AGPs | Notifiable organism but not notifiable disease |

| Suspected or confirmed Pathogen | Disease | Transmission based precautions (TBPs) required | Optimal placement while patient is considered infectious and until resolution of symptoms | Respiratory protection (RPE) for healthcare workers while patient is considered infectious ³ | Notifiable under Public Health Act 1984 and Health Protection Regulations 2010 ⁴ |
|--|---|---|---|--|---|
| Influenza virus (Endemic strains) | Influenza | Droplet | Single en-suite room | Fluid resistant surgical facemask (FRSM) for routine care and FFP3 or Hood for AGPs | Yes |
| Measles virus ² | Measles (rubeola) | Droplet/ Airborne | Isolation room/suite | FFP3 or Hood for routine care and AGPs | Yes |
| Met(h)icillin resistant Staphylococcus aureus (MRSA) | Colonisation, or clinical infection (skin and wound infections, endocarditis, pneumonia, osteomyelitis, urinary tract infections and bacteraemia) | Contact | Single en-suite room | FFP3 or Hood for AGPs only if pneumonia | No |
| Mumps virus ² | Mumps (infectious parotitis) | Droplet | Single en-suite room | Fluid resistant surgical facemask (FRSM) for routine care and FFP3 or Hood for AGPs | Yes |
| | Extrapulmonary Tuberculosis | Contact | Single en-suite room | FFP3 or Hood for AGPs | Yes |
| Mycobacterium tuberculosis complex | Pulmonary or laryngeal disease | Airborne | Isolation room/suite until patient has been established on appropriate antimicrobial treatment ⁵ and always if the patient has MDR or XDR TB | FFP3 or Hood for routine care and AGPs until patient has been established on appropriate antimicrobial treatment ⁵ and always if the patient has MDR or XDR TB | Yes |
| Mycoplasma pneumoniae | Pneumonia | Droplet | Single en-suite room | | |

| Suspected or confirmed Pathogen | Disease | Transmission based precautions (TBPs) required | Optimal placement while patient is considered infectious and until resolution of symptoms | Respiratory protection (RPE) for healthcare workers while patient is considered infectious ³ | Notifiable under Public Health Act 1984 and Health Protection Regulations 2010 ⁴ |
|--|---|---|--|--|---|
| Norovirus | Winter vomiting disease | Enteric | Single en-suite room | Fluid resistant surgical facemask (FRSM) if vomiting is present. | No (hospital outbreaks are reportable) |
| Panton Valentine Leukocidin (PVL) – positive Staphylococcus aureus | Skin and soft tissues infection, necrotising pneumonia, necrotising fasciitis, osteomyelitis, septic arthritis and pyomyositis, purpura fulminans | Contact | Single en-suite room | Fluid resistant surgical facemask (FRSM) for routine care and FFP3 or Hood for AGPs (only if pneumonia) | No |
| Parainfluenza virus¹ | Upper +/- lower respiratory tract infection | Droplet | Single en-suite room | Fluid resistant surgical facemask (FRSM) for routine care and FFP3 or Hood for AGPs | No |
| Parvovirus B19 – (Erythema infectiosum – Erythrovirus B19) | Slapped cheek syndrome | Droplet | Single en-suite room until the rash+/- arthralgia has developed | Fluid resistant surgical facemask (FRSM) for routine care and FFP3 or Hood for AGPs (Not required if the rash+/- arthralgia has developed) | No |
| Pneumocystis jirovecii | Pneumonia | Droplet | Single en-suite room in high risk settings eg ICU/PICU/NICU, oncology/haematology | Single en-suite room in high risk settings eg ICU/PICU/NICU, No requirement for RPE | |
| Pseudomonas aeruginosa | Pneumonia, bacteraemia, wound or surgical site infections, catheter- associated urinary tract infections, conjunctivitis in neonates | Droplet | Single en-suite room in high risk settings eg ICU/PICU/NICU, oncology/haematology | ingle en-suite room in high risk settings eg ICU/PICU/NICU, No requirement for RPE | |

| Suspected or confirmed Pathogen | Disease | Transmission based precautions (TBPs) required | Optimal placement while patient is considered infectious and until resolution of symptoms | Respiratory protection (RPE) for healthcare workers while patient is considered infectious ³ | Notifiable under Public Health Act 1984 and Health Protection Regulations 2010 ⁴ |
|--|--|--|--|---|---|
| Respiratory syncytial virus (RSV) ¹ | Upper +/- lower respiratory tract infection | Droplet | Single en-suite room | Fluid resistant surgical facemask (FRSM) for routine care and FFP3 or Hood for AGPs | No |
| Rotavirus | Gastroenteritis | Enteric | Single en-suite room | No requirement for RPE | No |
| Rubella virus ² | German Measles | Droplet | Single en-suite room | Fluid resistant surgical facemask (FRSM) for routine care and FFP3 or Hood for AGPs | Yes |
| Serratia marcescens | Pneumonia, bacteraemia, urinary tract infections, wound infections | Contact | Single en-suite room in high risk settings eg ICU/PICU/NICU, oncology/haematology | No requirement for RPE | No |
| Staphylococcus aureus (Enterotoxigenic) | Gastroenteritis, scalded skin syndrome | Contact - Enteric if the patient has diarrhoea | Single en-suite room (not required if lesions can be covered) | No requirement for RPE | No |
| Stenotrophomonas maltophilia | Bacteraemia, respiratory infections, urinary tract and surgical- site infections | Contact | Single en-suite room in high risk settings eg ICU/PICU/NICU, oncology/haematology | No requirement for RPE | No |
| Streptococcus pneumoniae | Pneumonia | Droplet | Single en-suite room (until patient has been established on appropriate antimicrobial treatment ⁵) | established on appropriate antimicrobial established on appropriate antimicrobial established on appropriate antimicrobial established on appropriate antimicrobial | |
| | Bacteraemia, meningitis, wound infection or infection in other normally sterile site | Contact | Single en-suite room (until patient has been established on appropriate antimicrobial treatment ⁵) | No requirement for RPE | Yes (presence in the wound is not notifiable) |

| Suspected or confirmed Pathogen | Disease | Transmission based precautions (TBPs) required | Optimal placement while patient is considered infectious and until resolution of symptoms | Respiratory protection (RPE) for healthcare workers while patient is considered infectious ³ | Notifiable under Public Health Act 1984 and Health Protection Regulations 2010 ⁴ |
|--|---|---|--|---|---|
| Streptococcus pyogenes | Respiratory infection | Droplet | Single en-suite room (until patient has been established on appropriate antimicrobial treatment ⁵) | Fluid resistant surgical facemask (FRSM) for routine care and FFP3 or Hood for AGPs until patient established on appropriate antimicrobial treatment ⁵ | No |
| (Group A Strep) | Invasive Group A strep Bacteraemia, meningitis, wound infection/infection in other normally sterile site | Contact | Single en-suite room (until patient has been established on appropriate antimicrobial treatment ⁵) | No requirement for RPE | Yes |
| Varicella virus ² See Herpes Zoster | Chickenpox | Airborne | Isolation room/suite | FFP3 or Hood for routine care and AGPs | Yes |
| Shiga-toxin producing Escherichia coli (STEC) Verocytotoxigenic Escherichia coli (including E.coli O157) Haemolytic uraemic syndrome (HUS) | Gastroenteritis, haemolytic uremic syndrome, thrombotic thrombocytopaenic purpura. | Contact - enteric if the patient has diarrhoea | Single en-suite room | No requirement for RPE | Some conditions notifiable (<u>refer to</u> guidance) |

Footnote 1

In routine clinical practice healthcare workers do not commonly wear masks when dealing with patients presenting with the "common cold" or "influenza – like illness". However, in a patient with undiagnosed respiratory illness where coughing and sneezing are significant features, or in the context of known widespread respiratory virus activity in the community or a suspected or confirmed outbreak of a respiratory illness in a closed or semi-closed setting, the need for appropriate respiratory and facial protection to be worn should be considered.

Footnote 2

In relation to childhood illnesses and use of RPE, no vaccine offers 100% protection and a small proportion of individuals acquire/become infected despite vaccination or known IgG immunity (previous infection). Vaccination is still the best protection against many infectious

diseases. If staff are uncertain of their immunisation status, they should discuss this with their occupational health provider. It is recommended that vaccinated individuals wear RPE as detailed in this appendix to minimise any residual risk, and to promote consistency in practice across all staff groups.

Footnote 3

The ocular route of transmission for pathogens spread by the droplet/airborne route while plausible lacks scientific evidence. This lack of evidence includes having very little certainty about what the incremental benefit of using eye protection routinely when using a FRSM/FFP3 respirator. Eye protection is considered to be necessary and worn if there is a risk of spraying or splashing of blood/body fluids from patient contact or procedure, and always when used with respirators during the performance of AGPs. This is line with published infection control quidance.

Footnote 4

Registered medical practitioners (RMPs) have a statutory duty to notify the 'proper officer' at their local council or local health protection team (HPT) of suspected cases of certain infectious diseases.

Complete a notification form immediately on diagnosis of a suspected notifiable disease. Don't wait for laboratory confirmation of a suspected infection or contamination before notification. Consult the UKHSA Notifiable Diseases poster (PDF, 1020KB, 1 page) for further information.

Send the form to the proper officer within 3 days, or notify them verbally within 24 hours if the case is urgent by phone, letter, encrypted email or secure fax machine.

If you need help, contact your local HPT using the postcode lookup. For more detail on reporting responsibilities of RMPs, see page 14 of Health Protection Legislation (England) Guidance 2010.

All proper officers must pass the entire notification to UKHSA within 3 days of a case being notified, or within 24 hours for urgent cases.

Footnote 5

Appropriate antimicrobial treatment will include the choice of treatment, dose, frequency and number of days of treatment. It will vary by organism and should be determined by the clinical team and informed by local and national prescribing guidance where available.

Footnote 6

Anthrax: during the bacteraemic phase split blood products should be removed immediately using sodium dichloroisocyanurate granules to prevent subsequent sporulation on contact with air / soil.

Appendix 11b Priority for Isolation

Isolation for Infection Prevention and Control purposes takes priority for the use of single rooms.

If any of the following are clinically suspected or confirmed by the laboratory the patient must be isolated. Contact Infection Prevention and Control and / or a Consultant Microbiologist for further information on managing these patients

| Organism / Infection | Isolation Category | Notifiable disease? | Additional Information |
|--|--------------------------------------|--|---|
| Monkeypox | Respiratory (Airborne) | Yes | Exclude immunocompromised or pregnant staff where possible |
| Drug resistant organism as advised by Microbiology | Contact | No | |
| Carbapenem producing Enterobacteriaceae (CPE) | Contact | No | |
| Chickenpox (varicella zoster virus) | Respiratory (Airborne) | Notifiable organism but not notifiable disease | Exclude non-immune and pregnant staff where possible. Isolate until lesions are dry and crusted. |
| Measles | Respiratory (Airborne) | Yes | Exclude non-immune and pregnant staff where possible. Infectious for 4 days before and 4 days after onset of rash |
| Middle Eastern Respiratory Syndrome Coronavirus (MERS CoV) | Respiratory (Airborne) | Yes | |
| SARS- CoV-2 (COVID-19) | Respiratory (Droplet or Airborne) | No | FFP3 masks or hoods required for Aerosol generating procedures. |
| Mycobacterium Tuberculosis (Pulmonary) | Respiratory (Airborne) | Yes | |
| Severe Acute Respiratory Syndrome (SARS) | Respiratory (Airborne) | Yes | |
| Viral Haemorrhagic Fevers, including Ebola | High Consequence Infection Disease | Yes | Contact IPCT / Consultant Microbiologist |
| Mumps | Respiratory (Droplet) | Yes | Exclude non-immune and pregnant staff where possible. |

The following patients require isolation. They are listed in order of priority for isolation (high to low)

| Organism / Infection | Isolation Category | Notifiable disease? | Inform Infection Prevention & Control | |
|---|---|--|---|--|
| Clostridioides difficile (Toxin positive and GDH positive toxin negative) | Enteric | No | Yes | Isolate until 72 hours of formed stool |
| Diarrhoea and/or vomiting of unknown cause | Enteric | Yes, if food poisoning | Yes | Collect stool sample |
| Epiglotittis or Supraglottitis (pending laboratory results) as these can be caused by Haemophilus influenzae type b or Streptococcus pyogenes (Group A Strep) | Contact (wound infection etc.) Respiratory (Droplet) | No | Only if Haemophilus influenzae type b or Streptococcus pyogenes (Group A Strep) | Isolate until patient established on appropriate antimicrobial treatment |
| Haemophilus influenzae type b causing any infection | Respiratory (Droplet) | Yes, if meningitis or bacteraemia | Yes | Isolate until patient established on appropriate antimicrobial treatment |
| MRSA in sputum, multiple sites, or skin shedders | Contact | No | Yes | Positive patients require decolonisation |
| E.coli 0157 | Enteric | Yes | Yes | Isolate until 48 hours symptom free |
| Shigella | Enteric | No | Yes | Isolate until 48 hours symptom free |
| Streptococcus pyogenes (Group A Strep) causing any infection | Contact | Yes – (invasive Group A strep) | Yes | Isolate until 48 hours after the start of effective antibiotic treatment. |
| Influenza | Respiratory (Droplet) | Yes | Yes | Isolate until clinical recovery |
| Neisseria meningitis causing meningitis or septicaemia | Respiratory (Droplet) | Yes | Yes | Isolate for first 24 hours of starting effective antibiotic treatment. |
| Whooping cough (Bordetella pertussis) | Respiratory (Droplet) | Yes | Yes | Isolate until 48 hours days of treatment or 21 days of start of symptoms if not treated |
| Respiratory Syncytial virus (RSV) | Respiratory (Droplet) | No | Yes | Isolate until clinical recovery |
| MRSA nasal only | contact | No | Yes | Positive patients require decolonisation |
| Herpes Zoster (Shingles) | Contact | Notifiable organism but not notifiable disease | Yes | Exclude non-immune and pregnant staff where possible. Isolate until lesions are dry and crusted. |
| Vancomycin resistant Enterococcus | Enteric | No | Yes | |
| Salmonella | Enteric | Yes | Yes | Isolate until symptom free |
| Acute viral encephalitis of all causes, | Respiratory | No | Yes | |

| including Herpes simplex virus | (Droplet) | | | |
|--|-----------------------|-----|-----|----------------------------------|
| Extended spectrum Beta Lactamase Contact | | No | Yes | |
| (ESBL) producing organisms | | | | |
| Amp C Beta Lactamase producing | Contact | No | Yes | |
| organisms | | | | |
| Common cold | Respiratory (Droplet) | No | No | |
| Body / Head Lice | Contact | No | No | |
| Scabies | Contact | No | Yes | Contact IPCT. Risk assessment is |
| | | | | required for patient and staff |
| | | | | treatment |
| Campylobacter | Enteric | Yes | No | |
| Cryptosporidium | Enteric | Yes | No | |

On patient admission, check Lorenzo and Meditech for IPC alerts to see if there has been an infection identified in the previous 12 months. If identified, please contact Infection Prevention and Control for advice regarding isolation.

Appendix 11c Bedside Precautions

It is recognised that, due to patient safety issues, it may not always be possible to isolate some patients. If a patient cannot be isolated the Infection Prevention and Control Team (IPCT) must be notified, and the reason documented.

Patient assessment must be reviewed at least daily and the patient isolated if safe to do so.

- Ensure the appropriate PPE (e.g. aprons / gloves/ and masks) is available
- Change all bins in the bay to infectious waste
- Equipment (BP cuff / tourniquet etc.) should be dedicated for the individual patient and cleaned or disposed of after use.
- ➤ Care must be taken when making beds to minimise dust dispersal i.e. fold sheets in on themselves do not shake.
- > Toilet facilities: Except for patients with diarrhoea/ enteric infections or those under respiratory precautions, the shared toilet in the bay may be used without additional actions providing:
 - o patients are supported with hand hygiene
 - blood or body fluid spillages are cleaned straight away, in line with the blood / body fluid spillage protocol
 - o the toilet is being routinely cleaned as per Trust standards
- The shared toilet may be used for patients under enteric and respiratory precautions providing:
 - it is cleaned by staff after each use including all the touch points toilet seat, taps, flush handle, door handle, nurse call bell etc.
 - A chlorine releasing agent should be used for those under enteric precautions. For other conditions cleaning after use using detergent is acceptable.
 - If you are unable to achieve this the patient will require their own commode.
 - If commodes are used, they may be stored in the ensuite but still must be cleaned after every National injection prevention and control manual for England use.
 - Visitors must be advised of any precautions they need to take

Appendix 11d Cohort bays for isolation purposes

Cohort bays may be advised by IPCT if the number of patients needing isolation for the same condition / symptoms exceeds the available side rooms, examples include norovirus and flu.

How to manage a cohort bay:

- All unnecessary equipment should be cleaned and removed from the bay.
- > Ensure the appropriate PPE (e.g. aprons / gloves/ and masks) is available
- Change all bins in the bay to infectious waste
- Dedicate equipment for the bay, ideally per patient. Shared equipment must be cleaned between patients.
- > PPE must be changed between each patient in the bay and hand hygiene performed
- PPE should be removed in the bay then hands should be washed at the bay sink before leaving. If handling the door on the way out (e.g. Derby door) then also use alcohol hand sanitiser after closing the door
- ➤ Ideally, a cohort bay should have doors that close, in the absence of doors; the 'Derby Door' should be used wherever possible. This will not be appropriate for all patient groups and a risk assessment should be undertaken
- > Patients and visitors must be informed of the need for cohort nursing and the precautions they should take.
- Visitors are not required to wear PPE unless giving personal care. In this circumstance PPE should be offered under the same guidance of staff according to reason for the cohort

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Appendix 11e Aide memoire for optimal patient placement and respiratory protective equipment (RPE) for high consequence infectious diseases

High Consequence Infectious Diseases

- Should be discussed immediately with specialist teams including health protection teams and the imported fever service. UKHSA which provides 24-hour, 7 days a week telephone access to expert clinical and microbiological advice. Hospital doctors across the UK can contact the IFS after discussion with the local microbiology, virology or infectious disease consultant.
- Once an HCID has been confirmed cases in England should be transferred rapidly to a designated HCID Treatment Centre.

HCID is defined according to the following criteria:

- acute infectious disease
- typically has a high case-fatality rate
- may not have effective prophylaxis or treatment
- often difficult to recognise and detect rapidly
- ability to spread in the community and within healthcare settings
- requires an enhanced individual, population and system response to ensure it is managed effectively, efficiently and safely

HCIDs are further divided into contact and airborne groups. Contact HCIDS are usually spread by direct contact with an infected patient or infected fluids, tissues and other materials, or by indirect contact with contaminated material and fomites. Airborne HCIDS are spread by respiratory droplets or aerosol transmission, in addition to contact routes of transmission. Specific infection prevention and control (IPC) measures are required for suspected and confirmed HCID cases, in all healthcare settings.

| HCID | Pathogen | Predominant mechanism of spread | Optimal Placement while suspected cases | Required PPE | Notifiable under Public Health Act 1984 and Health Protection Regulations 2010. |
|--------------|--|---------------------------------|--|--------------|--|
| Contact HCID | Argentine haemorrhagic fever (Junin virus) Bolivian haemorrhagic fever (Machupo virus) Crimean Congo haemorrhagic fever (CCHF) Ebola virus disease (EVD) Lassa Fever Lujo virus disease Marburg virus disease (MVD) Severe fever with thrombocytopaenia syndrome (SFTS). | Contact | Complete risk assessment to define possibility of HCID in suspected cases Single side room with en- suite facilities / dedicated commode. | · | Yes |

| HCID | Pathogen | Predominant mechanism of spread | Optimal Placement while suspected cases | Required PPE | Notifiable under Public Health Act 1984 and Health Protection Regulations 2010. |
|---------------|---|--|--|---|--|
| Airborne HCID | Andes virus infection (hantavirus) Avian influenza A H7N9 and H5N1 Avian influenza A H5N6 and H7N7 Middle East respiratory syndrome (MERS) mpox (Monkeypox)¹ Niapah virus infection Pneumonic plague (Yersinia pestis) Severe acute respiratory syndrome (SARS). | Unknown; assume airborne until further information available | Single side room with ensuite, ideally under negative pressure | As a minimum FFP3 respirator, gown, gloves and eye protection Additional contact measures may be required according to pathogen: See above for high possibility contact HCID | Yes |

1. The Advisory Committee on Dangerous Pathogens (ACDP) has recommended that all of Clade II MPXV (formerly West African clade) should no longer be classified as an HCID. Clade I (formerly known as Central African or Congo basin clade) should remain an HCID. The 4 nations public health agencies have reviewed this advice and agreed with the view of ACDP. Refer to UKHSA principles for control of monkeypox in England (May 2022) for guidance on management of the strain of monkeypox virus (MPXV) currently in community transmission within the UK (Clade IIb, B.1 lineage).

Further resources:

High consequence infectious diseases (HCID) - GOV.UK (www.gov.uk)

Viral haemorrhagic fever: ACDP algorithm and guidance on management of patients - GOV.UK (www.gov.uk)

VHF_Algo.pdf (publishing.service.gov.uk)

Appendix 12: Transmission based precautions for deceased patients with infection

As per section 2.6 of the NIPCM, the principles of SICPs and TBPs continue to apply while deceased individuals remain in the care environment. This is due to the ongoing risk of infectious transmission via contact although the risk is usually lower than for living patients. Additional precautions may be required depending on the organism and activities carried out (see table).

| Infection | Causative agent | Hazard Group | Is a body bag needed ¹ ? | Can the body be viewed? | Can post mortem be carried out? ² | Can hygienic treatment be carried out? ³ | Can embalming be carried out? ² | | |
|---|---|-----------------|--|-------------------------|--|---|---|--|--|
| Airborne: small p | Airborne: small particles that can remain airborne with potential for transmission by inhalation | | | | | | | | |
| Plague (Pneumonic and bubonic) | Yersinia pestis | 3 | Yes | Yes | If an appropria te facility is found | Consult specialis t advice | Consult specialist advice | | |
| Tuberculosis | Mycobacteriu m tuberculosis | 3 | Yes | Yes | Yes | Yes | Yes | | |
| Middle Eastern Respiratory Syndrome (MERS) | MERS coronavirus | 3 | Yes | Yes | Yes | Yes | Yes | | |
| Severe acute respiratory syndromes | eg SARS coronavirus see HSE Handling the deceased with suspected or confirmed COVID-19 - HSE | 3 | Yes | Yes | Yes | Yes | Yes | | |
| | Droplet : large particles that do not remain airborne for very long and do not travel far from source with potential for transmission via mucocutaneous routes (ie mouth, nose, or eyes) | | | | | | | | |
| Meningococcal septicaemia (Meningitis) | Neisseria meningitidis | 2 | No | Yes | Yes | Yes | Yes | | |
| Non- meningococcal meningitis | Various bacteria including Haemophilus influenzae and also viruses | - | No | Yes | Yes | Yes | Yes | | |
| Influenza (animal origin) | eg H5 and H7 influenza viruses | 3 | No | Yes | Yes | Yes | Yes | | |
| Diphtheria | Corynebacteri um diphtheriae | 2 | No | Yes | Yes | Yes | Yes | | |

| Infection | Causative agent | Hazard Group | Is a body bag needed ¹ ? | Can the body be viewed? | Can post mortem be carried out? ² | Can hygienic treatment be carried out? ³ | Can embalming be carried out? ² | |
|---|---|-----------------|--|-------------------------|--|---|---|--|
| Contact: either direct via hands of employees, or indirect via equipment and other contaminated articles where transmission is primarily via an ingestion route | | | | | | | | |
| Invasive streptococcal infection | Streptococcus pyogenes (Group A) | 2 | Yes | Yes | Yes | No | No | |
| Dysentery (shigellosis) | Shigella dysenteriae (type 1) | 3 | Advised | Yes | Yes | Yes | Yes | |
| Methicillin- resistant Staphylococcus aureus (MRSA) | Methicillin- resistant Staphylococcu s aureus | 2 | No | Yes | Yes | Yes | Yes | |
| Hepatitis A | Hepatitis A virus | 2 | No | Yes | Yes | Yes | Yes | |
| Hepatitis E | Hepatitis E virus | 3 | No | Yes | Yes | Yes | Yes | |
| Enteric fever (typhoid/para typhoid) | Salmonella typhi/paratyphi | 3 | Advised | Yes | Yes | Yes | Yes | |
| Brucellosis | Brucella melitensis, B. arbortus, B. suis | 3 | No | Yes | Yes | Yes | Yes | |
| Haemolytic uraemic syndrome | Verocytotoxin/ shiga toxin producing <i>E.coli</i> (eg O157:H7) | 3 | No | Yes | Yes | Yes | Yes | |
| | irect or indirect co or via broken skird mouth | | | | | | | |
| Acquired Immune Deficiency Syndrome related illness | Human immune- deficiency virus | 3 | No | Yes | Yes | Yes | Yes | |
| Anthrax | Bacillus anthracis | 3 | Yes | No | Yes⁴ | No | No | |
| Hepatitis B, D and C | Hepatitis B, D and C viruses | 3 | No | Yes | Yes | Yes | Yes | |
| Rabies | Lyssaviruses | 3 | No | Yes | No | No | No | |
| Viral haemorrhagic fevers | See appendix 11b | 4 | Yes⁵ | No | No | No | No | |
| Contact: either direct or indirect contact with body fluids (eg brain and other neurological tissue) via a skin-penetrating injury or via broken skin | | | | | | | | |
| Transmissible spongiform encephalopathi es (eg vCJD) | Various prions | 3 | Yes | Yes | Yes | Yes | No | |

Notes

- It is advised that a body bag is used for the deceased in all cases where there is (or is likely to be) leakage of bodily
- When carrying out higher risk procedures such as post-mortem or embalming, consideration should be given to the need for additional measures to prevent contamination of equipment

| Infection Causa | tive agent Hazard Group | Is a body bag needed ¹ ? | Can the body be viewed? | Can post mortem be carried out? ² | Can hygienic treatment be carried out? ³ | Can embalming be carried out? ² |
|-----------------|----------------------------|--|-------------------------|--|---|---|
|-----------------|----------------------------|--|-------------------------|--|---|---|

and the environment and to prevent staff exposure to infectious material eg through additional PPE and use of safer sharps devices.

- Hygienic treatment refers to washing and/or dressing of the deceased.
- ⁴ Where anthrax infection is suspected, before undertaking a post mortem the rationale for the procedure should be carefully considered; particularly where examination may increase the potential for aerosol generation.
- A double body bag must be used.

NB Hazard group 4 and HCID will be transported by HART teams (see section 2.6)